

Automated Homology-based Approach for the Identification of Transposable Elements: TESeeker

Ryan C. Kennedy
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Transposable Elements (TEs)

- ❑ First found and analyzed by Barbara McClintock in 1948
 - Won Nobel Prize in 1983
- ❑ TEs are mobile pieces of DNA
- ❑ Typically divided into Class I and Class II elements
 - Class I elements are RNA-mediated
 - Class II elements are DNA-mediated
- ❑ Example *mariner* Class II TE:



TA	ACGC...GTAA	GTATCAGCCA...CAAATTACG	TTAC...GCGT	TA
Target Site Duplication	Inverted Repeat	Transposase	Inverted Repeat	Target Site Duplication
2 bp	20-30 bp	~900 bp	20-30 bp	2 bp

Motivation

□ Why study transposable elements?

- Have been found in all eukaryotic genomes
- Occupy large portions of genomes
 - 50% of human genome
 - 47% of *Aedes aegypti* mosquito genome
- Can influence genome evolution and gene expression

■ Mouse Genome Sequencing Consortium:

“The single most prevalent feature of mammalian genomes is their repetitive sequences, most of which are interspersed repeats representing ‘fossils’ of transposable elements. *Transposable elements are a principal force in reshaping the genome, and their fossils thus provide powerful reporters for measuring evolutionary forces acting on the genome.*”

R.H. Waterston, et al. Initial sequencing and comparative analysis of the mouse genome. *Nature*, 420:520-562, December 2002.

TE Discovery Techniques

- ❑ Bergman and Quesneville categorize TE discovery into four categories:
 - 1) Comparative Genomic Methods
 - ❑ Perform multiple sequence alignment of related genomes and look for large changes amongst them
 - ❑ Good for finding new TE families, but relies on readily available, properly sequenced, related genomes
 - 2) *De novo*
 - ❑ Detect similar sequences found throughout the genome and cluster
 - ❑ Can discover new TE families, but often difficult to distinguish closely related TEs

TE Discovery Techniques

3) Structure-based

- ❑ Use TE structural data, such as inverted repeats, to find TEs
- ❑ Works well for characterized TEs, but does not locate degraded TEs or TEs with non-distinct structures

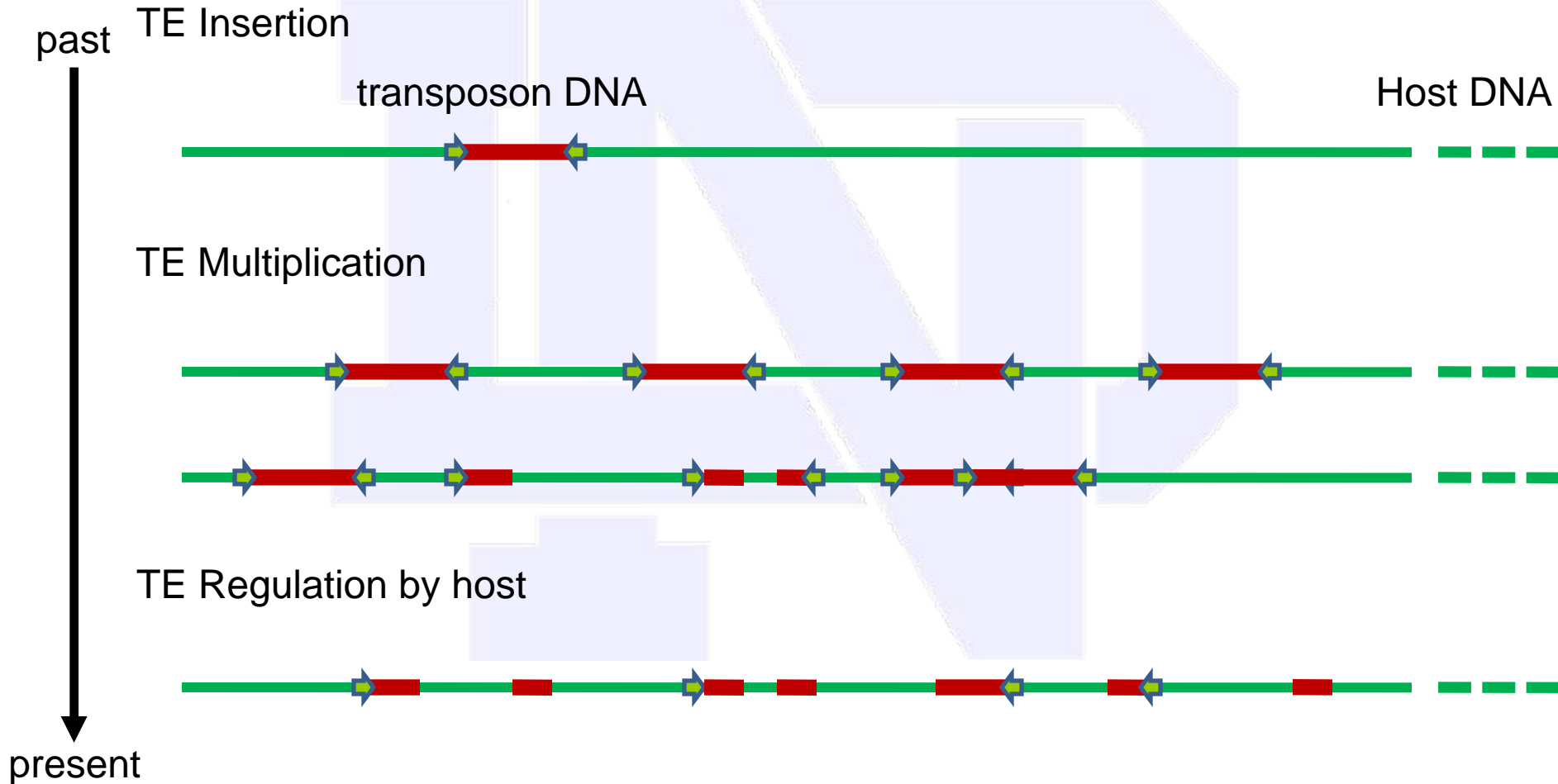
4) Homology-based (our approach)

- ❑ Use known TEs as seeds to search in novel genomes
- ❑ Can discover new TE families, but requires additional verification

Challenges in Locating TEs

- Although present in all eukaryotic genomes, difficult to annotate
 - Varying structural characteristics
 - Mobile nature often leads to copies within copies
 - TEs often are very degraded

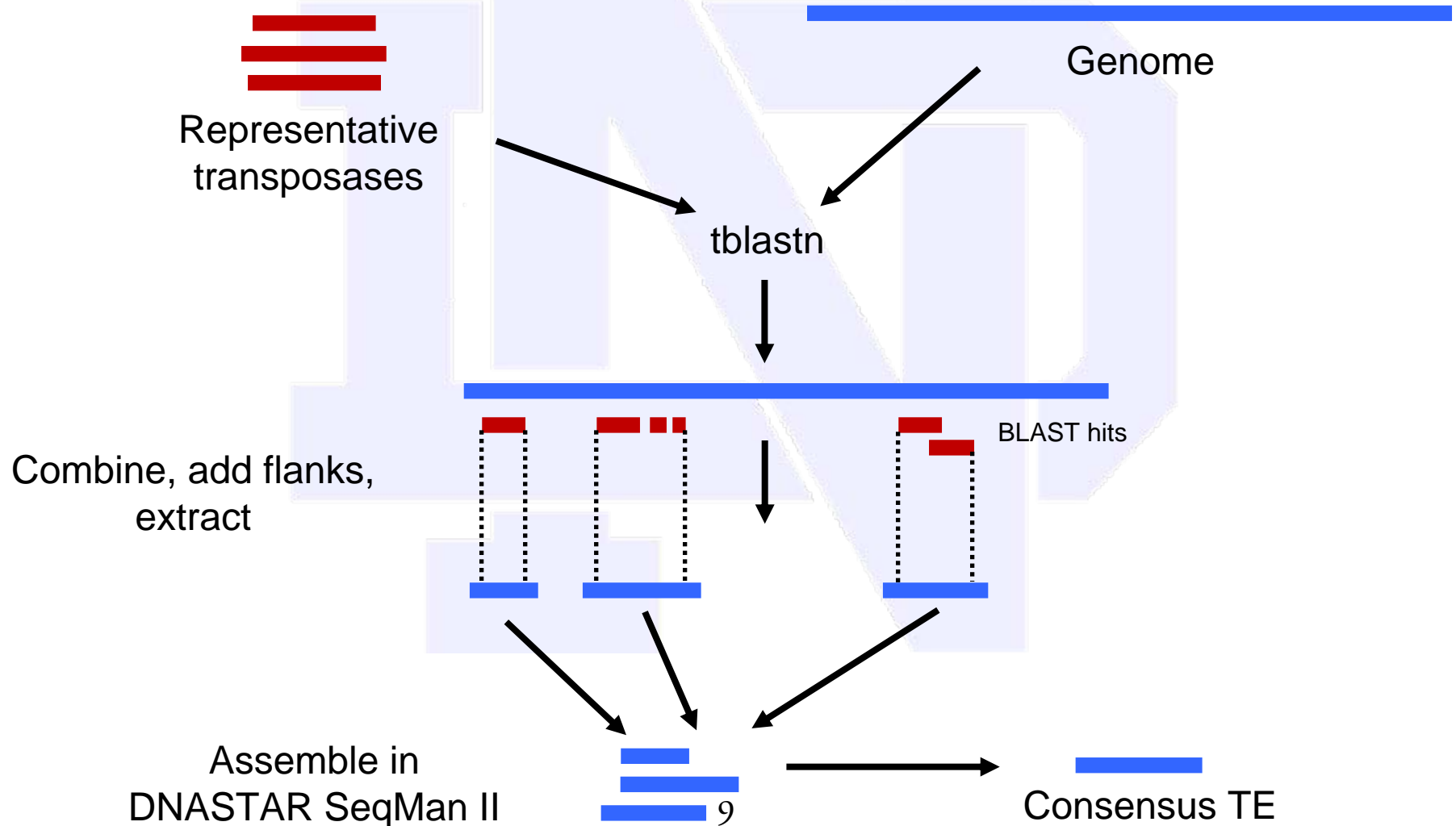
Class II TE Evolution



Manual Approach

- ❑ Developed and utilized during TE search on very different genome projects:
 - *Pediculus humanus humanus* (body louse)
 - ❑ Comprehensive search for all TE families
 - *Culex quinquefasciatus* (mosquito)
 - ❑ Search for non-LTR TEs
- ❑ Homology-based
 - Assembled representative TE library of high-quality TEs
 - ❑ Intact open reading frames
 - Results appear in TE sections of respective genome papers

Manual Approach



P. humanus humanus Results



Class I	Family	Element	Length (bp)	Full-length Copies	Partial Hits	Density
Non-LTR	SART	<i>Hope-like</i>	4655	1	522	0.18%
	R4	<i>Dong-like</i>	5266	4	1739	0.45%
LTR	Ty3/gypsy	<i>Mdg1</i>	5395	2	976	0.28%
Class II	Family	Element	Length (bp)	Full-length Copies	Copies	Density
	Mariner/Tc1	<i>mariner</i>	1276	24	216	0.09%
TOTAL						1.0%

E.F. Kirkness et al., "Genome sequences of the human body louse and its primary endosymbiont provide insights into the permanent parasitic lifestyle." *Proceedings of the National Academy of Sciences*, 107(27):12168-12173, July 2010.

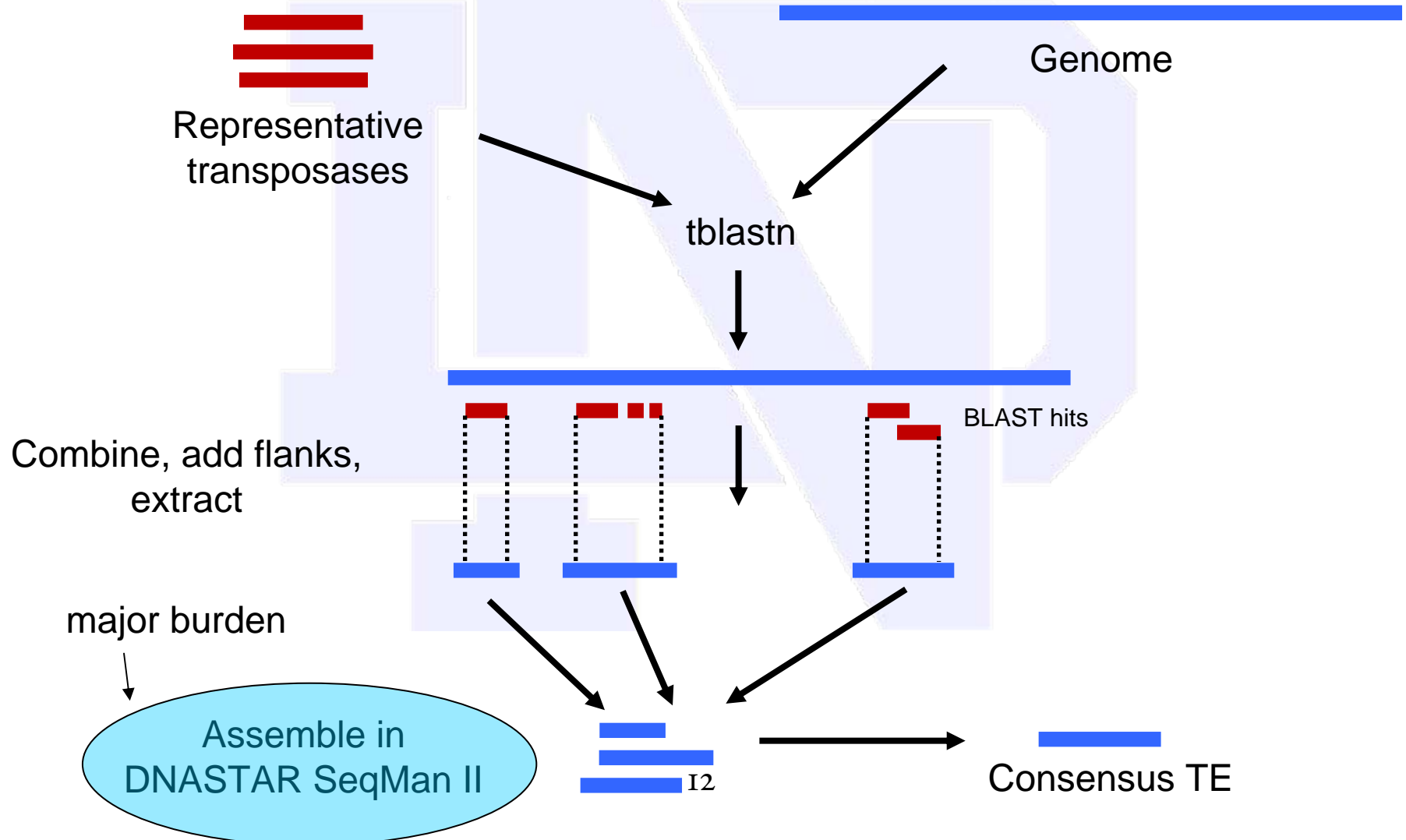
C. quinquefasciatus Results



Class I	Family	Full-length Copies	Partial Hits	Density
Non-LTR	CR1	31	973	0.28%
	I	11	63	0.02%
	Jockey	14	5028	1.77%
	L1	57	662	0.15%
	L2	9	1416	0.61%
	Loa	9	184	0.09%
	Loner	2	127	0.12%
	Outcast	4	15	0.00%
	R1	32	250	0.14%
	RTE	8	892	0.38%
	Unclassified LINE	32	11,117	0.88%
TOTAL				4.44%

P. Arensburger et al., "Sequence of *Culex quinquefasciatus* Establishes a Platform for vector Mosquito Comparative Genomics." *Science*, 330(6000):86-88, October 2010.

Manual Approach



DNAStar SeqMan II

- ❑ Manually trimming hits and additional processing is time consuming
- ❑ Can only assemble limited number of sequences at a time

The screenshot displays the 'Alignment of Contig 86' window in DNAStar SeqMan II. The window title is 'Alignment of Contig 86'. The position is 737, and the total length is 4.193kb. The alignment shows multiple sequences (1103172108213-8527 to 1103172107855-8570) aligned to a reference sequence. The reference sequence is ATATTTT-CGTTA-AAT-AATTTG--TTTT-A-T-GTA-AAT---TATTGGGTTGGCMAATAAGTAACTGCGSATTTTACCAACAGATAGTT. The sequences are color-coded: red for matches, green for mismatches, and blue for gaps. The alignment is shown in a grid format with columns for each sequence. The sequences are listed on the left, and the alignment is shown on the right. The alignment is shown in a grid format with columns for each sequence. The sequences are listed on the left, and the alignment is shown on the right. The alignment is shown in a grid format with columns for each sequence. The sequences are listed on the left, and the alignment is shown on the right.

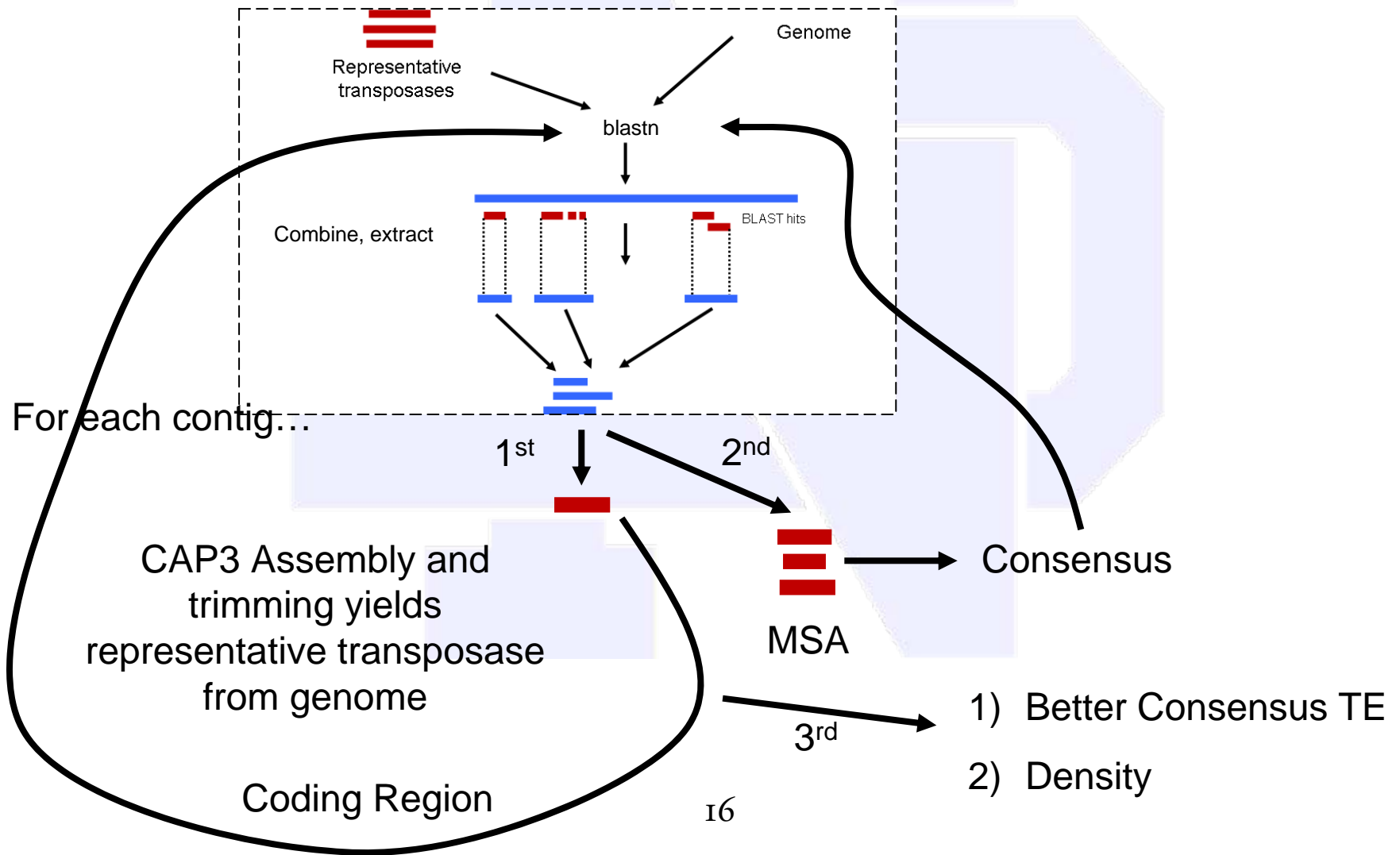
Automated Approach

- ❑ Homology-based
- ❑ Replace DNASTAR SeqMan II and manual analysis with other tools
 - CAP3, Clustal, various scripts
- ❑ Iterative- repeat steps if necessary

Automated Approach Steps

1. Identify transposase in target genome
 2. Find copies in target genome with flanks
 3. Generate consensus from multiple sequence alignment of copies
 4. Use consensus to identify TE
-
- Output: putative high-quality consensus TE which can in turn be used locate instances within the genome
 - Runs in a matter of minutes/hours
 - Dependent on genome size, size of representative TEs, and richness of TEs in the genome
 - Runs via web interface or via automated scripts

Automated Approach



Step 1

- Identify transposase(s) in target genome
 - *tblastn* representative transposases against genome
 - Parse BLAST file with the following parameters:
 - combine threshold: maximum distance sequences can be apart to join as a single hit
 - minimum length percentage: must be at least this percentage of query sequence to be considered
 - e-value cutoff: ignore everything worse than this value, typically 1E-20
 - flank size: amount of extra sequence to add to each end of hit (0)
 - Extract genomic sequences from above and iteratively assemble with CAP3
 - With CAP3, specify quality window size and threshold, as well as combine threshold



transposase(s) within genome

Step 2

- Find copies in target genome with flanks
 - *blastn* transposase(s) against genome
 - Parse BLAST file with the following parameters:
 - combine threshold: maximum distance sequences can be apart to join as a single hit
 - minimum length percentage: must be at least this percentage of query sequence to be considered
 - e-value cutoff: ignore everything worse than this value, typically 1E-20
 - flank size: amount of extra sequence to add to each end of hit
 - Extract genomic sequences from above



Copies within genome with flanks

Step 3

- ❑ Obtain consensus from multiple sequence alignment (MSA) of copies
 - Perform MSA on sequences
 - Generate consensus from MSA
 - ❑ Can specify percentage of nucleotides that must be common amongst sequences to count in consensus



Putative Consensus

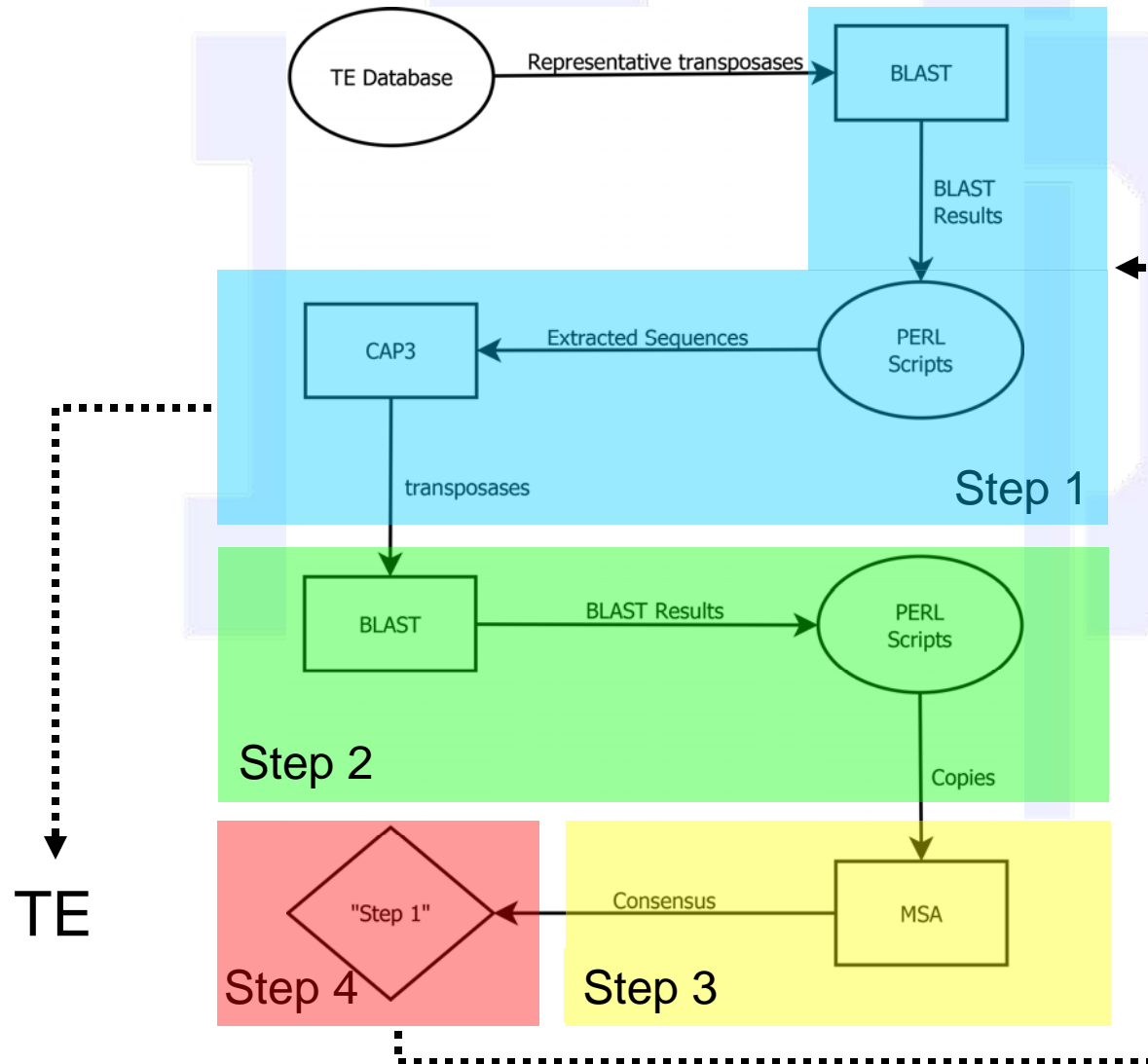
Step 4

- Use consensus to identify proper TE
 - *blastn* representative transposases against genome
 - Parse BLAST file with the following parameters:
 - combine threshold: maximum distance sequences can be apart to join as a single hit
 - minimum length percentage: must be at least this percentage of query sequence to be considered
 - e-value cutoff: ignore everything worse than this value, typically 1E-20
 - flank size: amount of extra sequence to add to each end of hit
 - Extract genomic sequences from above and iteratively assemble with CAP3
 - With CAP3, specify quality window size and threshold, as well as combine threshold



Consensus TE → Density

Automated Approach Schematic

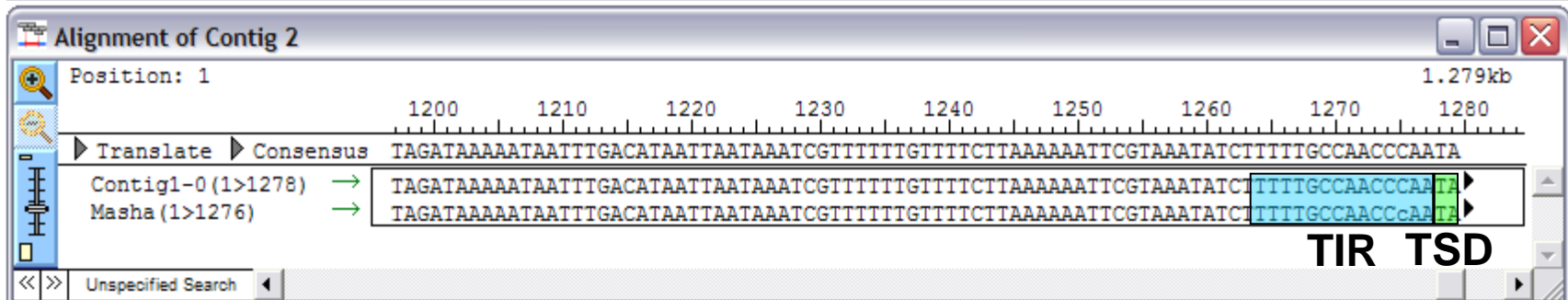
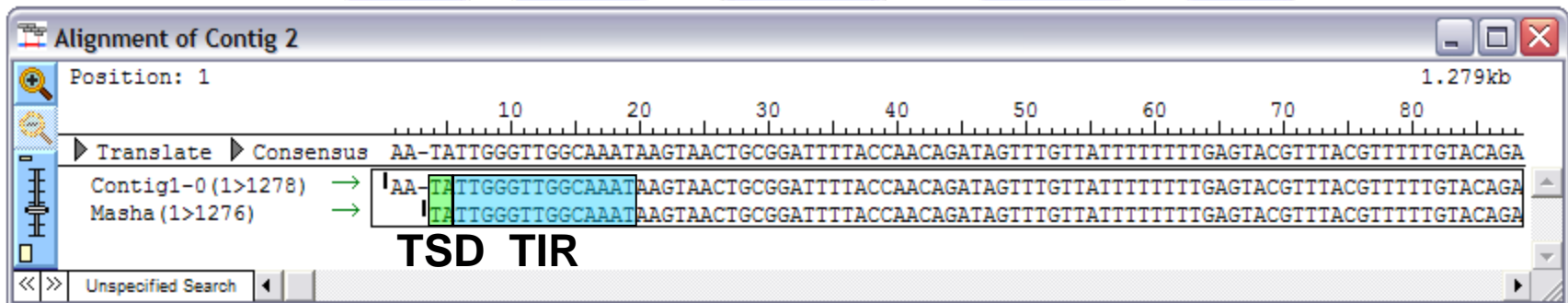


Validation Strategy

1. Initially evaluated automated approach on *P. humanus* and *C. quinquefasciatus*
 - Validate against high-quality manually verified annotation
 - Identify default starting parameters
2. Check automated results versus published results
3. Genomes in General:
 - Translate consensus TE sequences
 - Identify open reading frame
 - *blastp* against *non-redundant protein (nr)* database at NCBI and check for conserved domains/hits
 - Can check for structural signatures

Validation (1): *P. humanus humanus mariner*

- ❑ Full *mariner* element identified following Step 4
- ❑ Validated against manual effort
 - TSDs; 14 bp terminal inverted repeats (TIRs); well-trimmed



Validation (2): *Anopheles gambiae* PEST

□ *P* elements (Class II)

- Sarkar et al. (2003) identified 6 distinct elements
- Oliveira de Carvalho et al. (2004) identified 4 additional elements
- Quesneville et al. (2006) identified 9 elements at least 30% divergent at nucleotide level
- **Total:** 12 elements at least 30% divergent at nucleotide level
- Automated Approach
 - Identified 11/12 elements + 2 partial hits
 - Captured TIRs where previously described

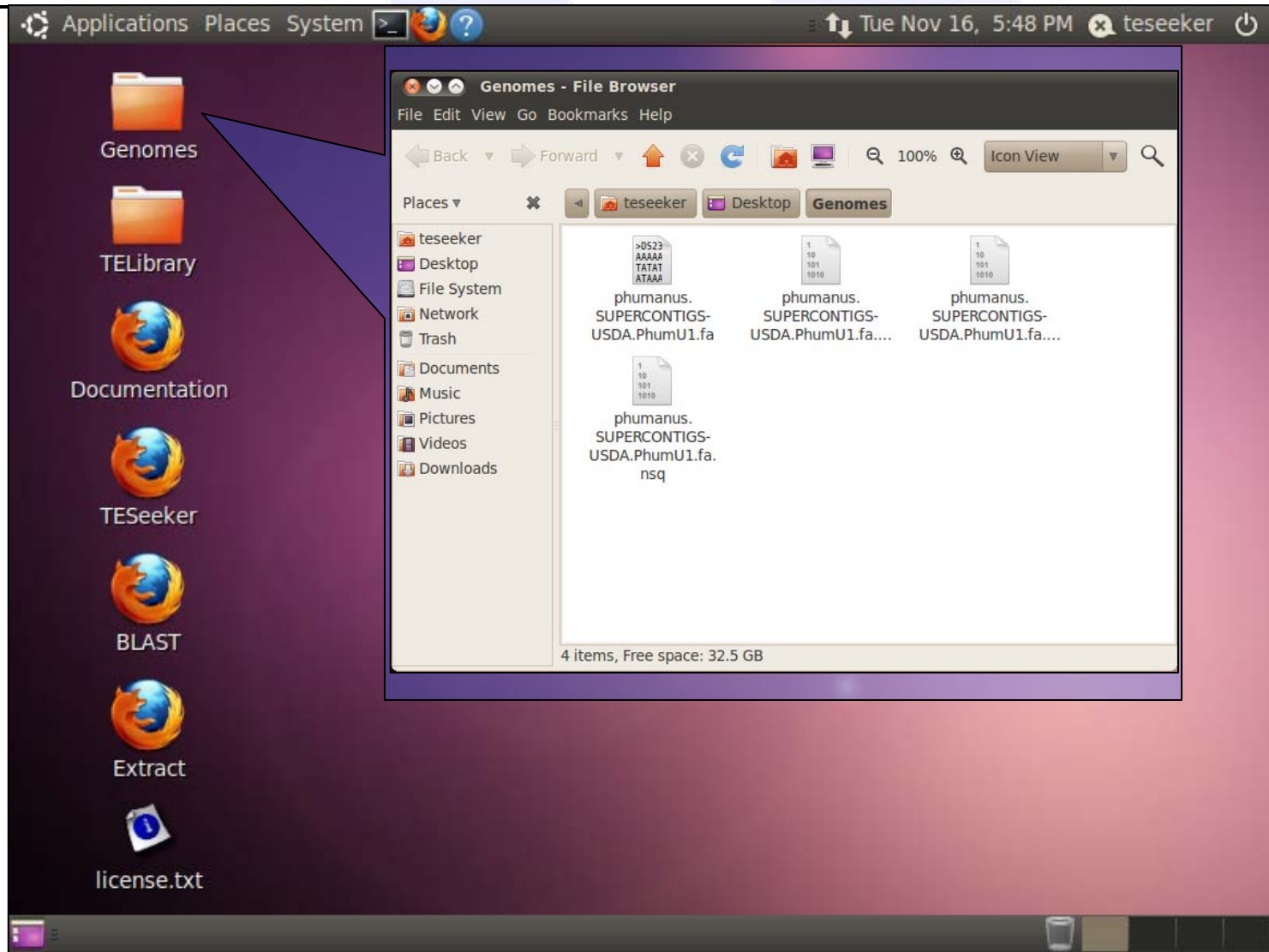
Validation (3)

- Searched for *mariner* in a number of genomes
 - In agreement where previously reported
 - Human, frog, chicken
 - In agreement where not reported
 - Dog, cat, horse
- Possible discovery
 - *Drosophila melanogaster* putative *mariner*
 - 1061 bp element has TIRs
 - 26 bp TIRs
 - no apparent TSDs
 - Single full-length copy, as well as several partial hits
 - Transposase is most similar to that of *Chymomyza amoena*, 77% identical at the amino acid level
 - Searches for this element in existing TE annotations for *D. melanogaster* produced no hits

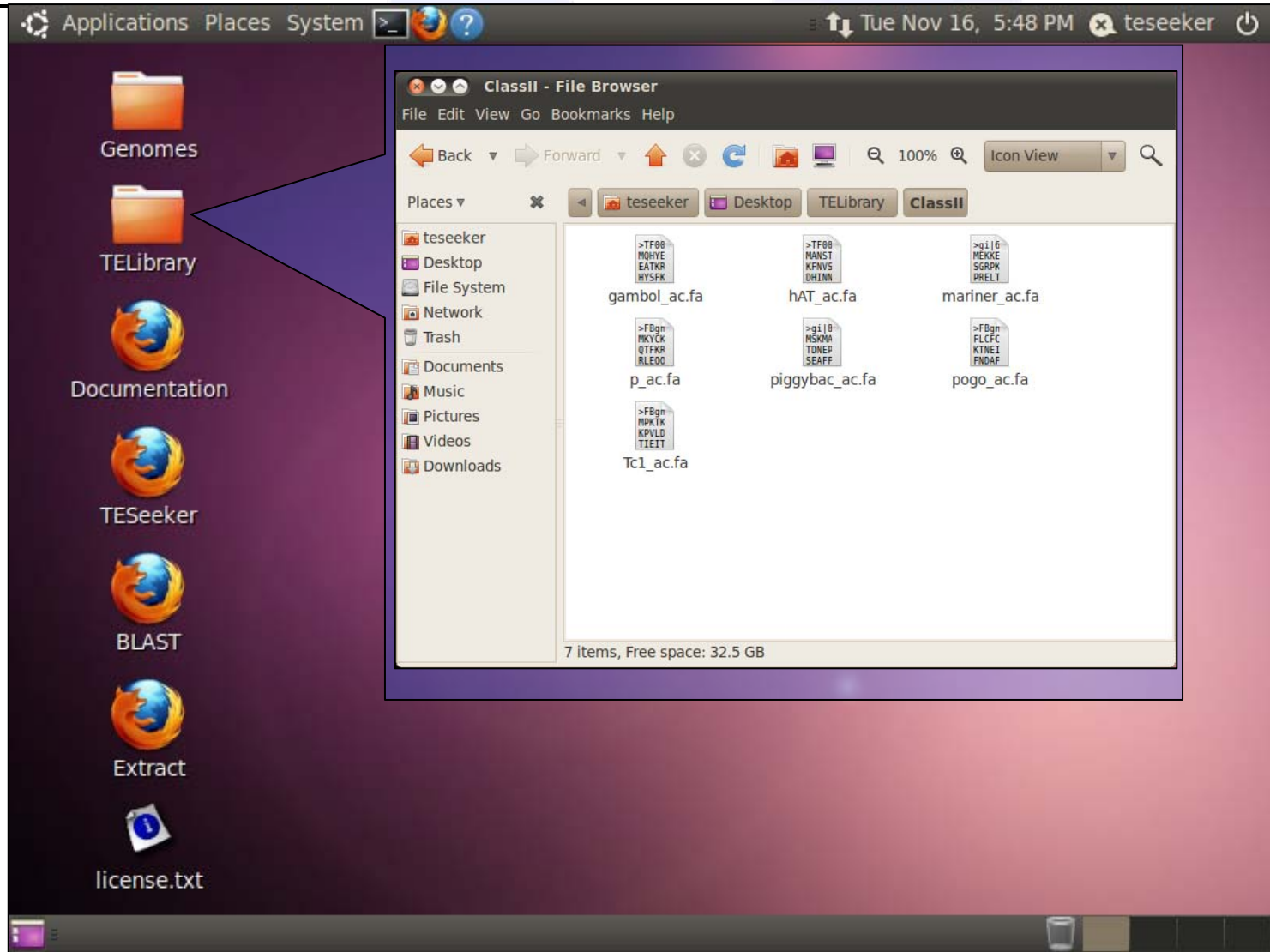
Implementation

- ❑ Approach implemented as TESeeker
 - VirtualBox virtual appliance
 - ❑ Cross-platform
 - ❑ Completely configured, no need to install scripts
 - Provide only genome FASTA file
 - Optionally provide additional library files
 - ❑ Local web interface
- ❑ <http://www.nd.edu/~tseeker>
 - Virtual appliance
 - Documentation
 - TE Library

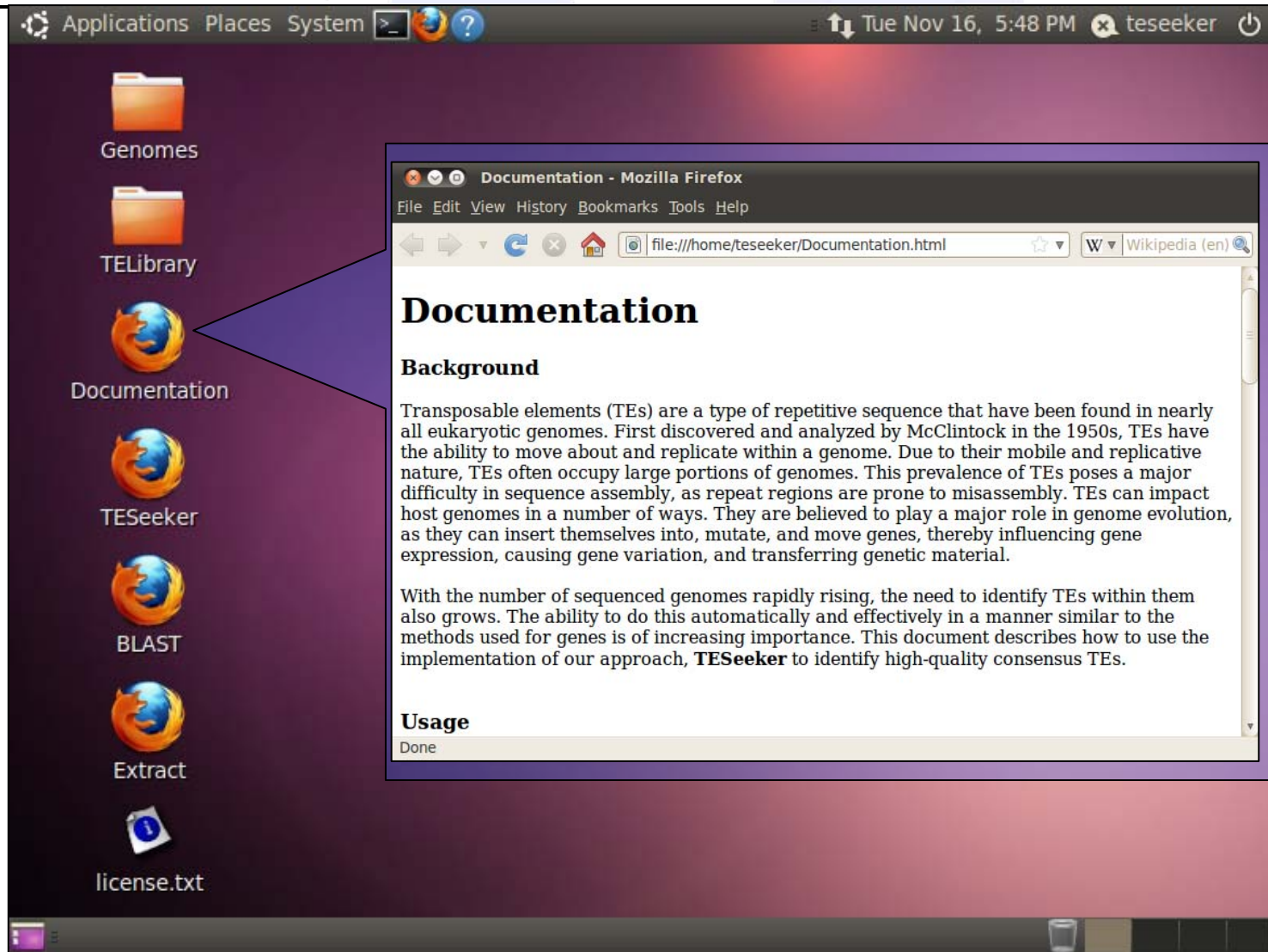
TESeeker Desktop



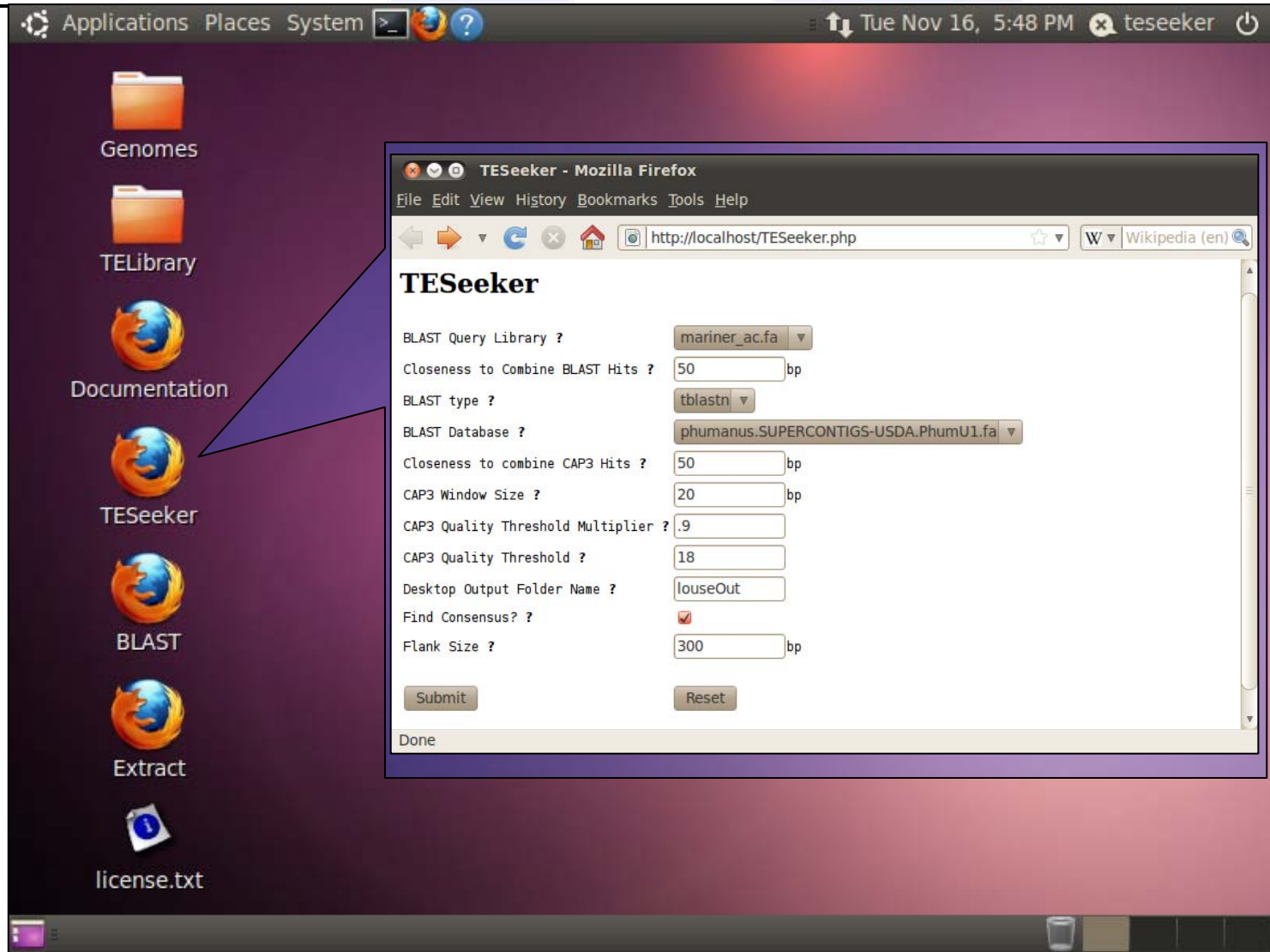
TESeeker Desktop



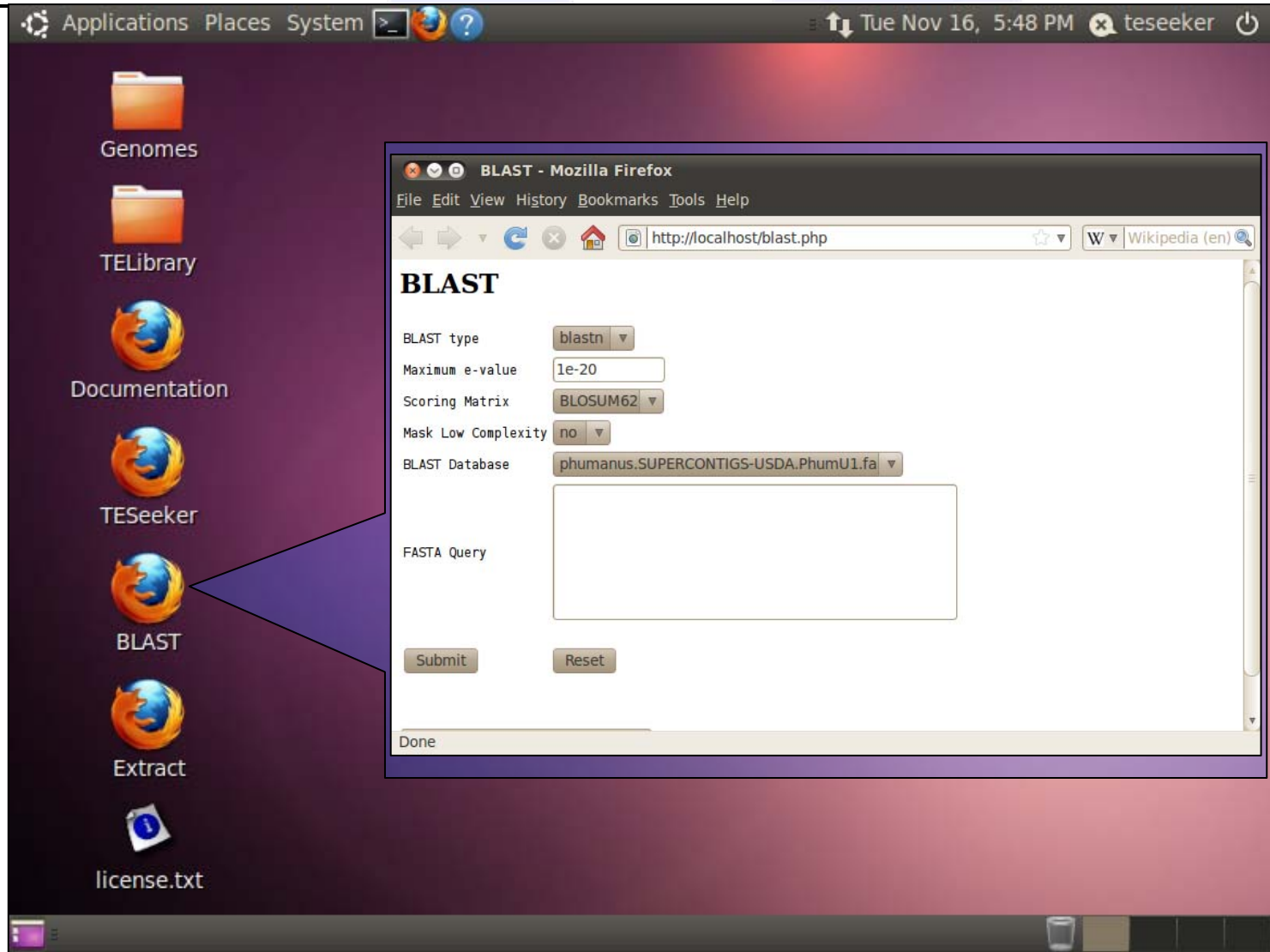
TESeeker Desktop



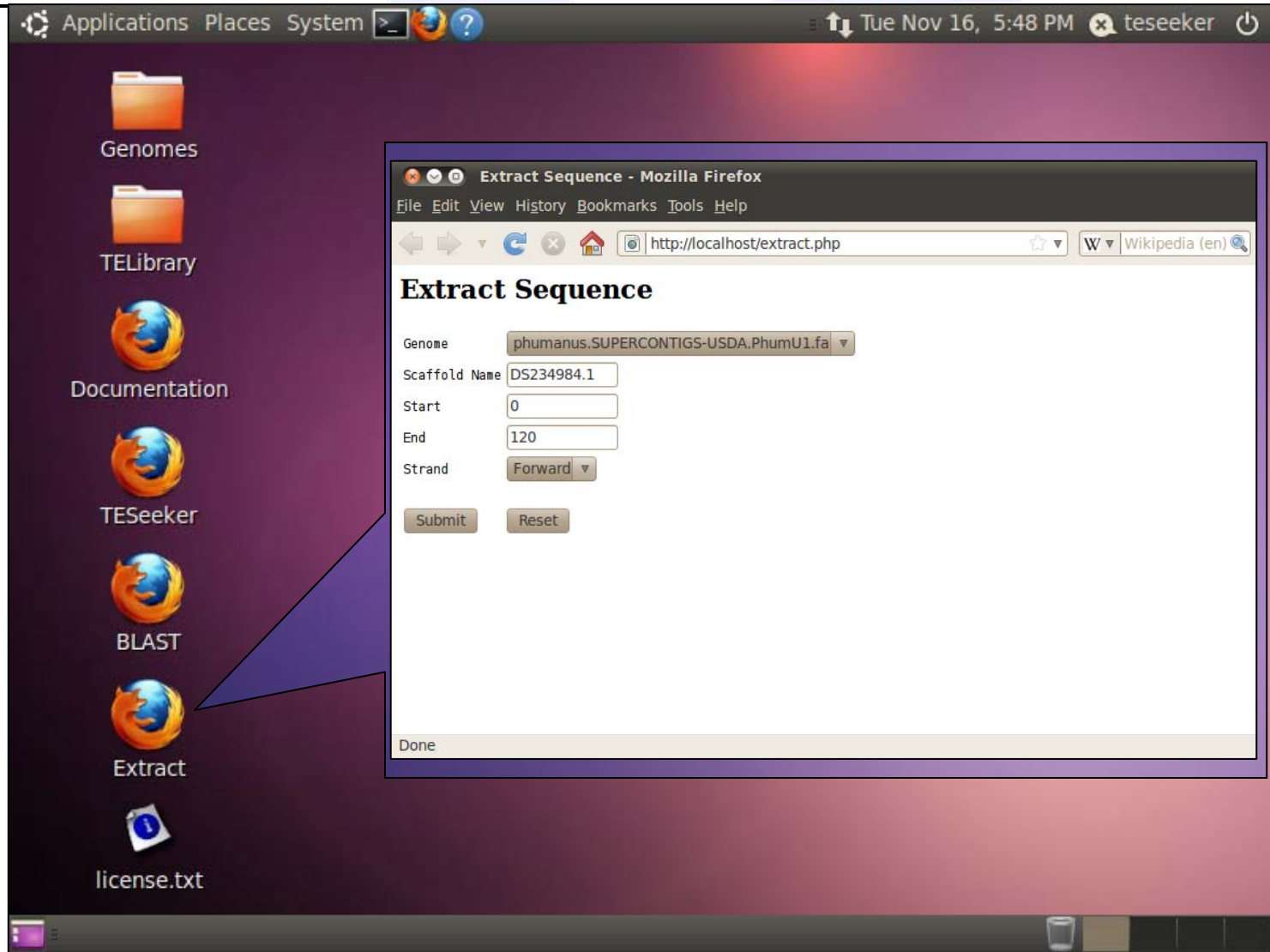
TESeeker Desktop



TESeeker Desktop



TESeeker Desktop



TESeeker Walkthrough

- ❑ Identify *mariner* element in *P. humanus humanus*:
 - Start with default parameters
 - Make sure genome file and library file are present
 - Start search

TESeeker Desktop



TESeeker Local Web Interface

TESeeker - Mozilla Firefox

File Edit View History Bookmarks Tools Help

http://localhost/TESeeker.php

Wikipedia (en)

TESeeker

BLAST Query Library

Closeness to Combine BLAST Hits bp

BLAST type

BLAST Database

Closeness to combine CAP3 Hits bp

CAP3 Window Size bp

CAP3 Quality Threshold Multiplier

CAP3 Quality Threshold

Desktop Output Folder Name

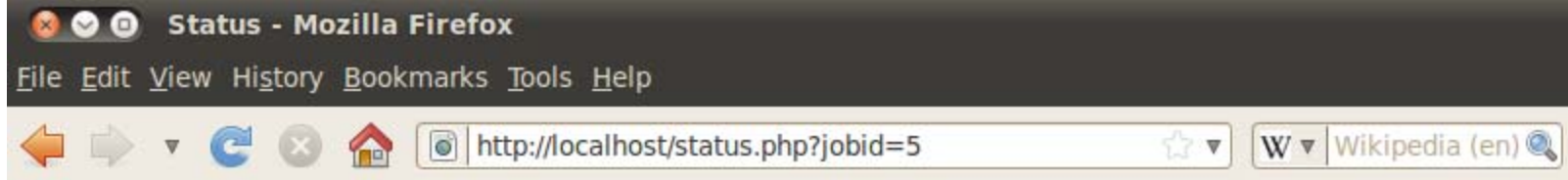
Find Consensus? ☒

Flank Size bp

Done

Y OF ME

TESeeker Status



Job 5 is Running

TESeeker Status



TESeeker Results

Index of file:///home/teseeker/Desktop/louseOut - Mozilla Firefox

File Edit View History Bookmarks Tools Help

file:///home/teseeker/Desktop/louseOut

Wikipedia (en)

Index of file:///home/teseeker/Desktop/louseOut

[Up to higher level directory](#)

Name	Size	Last Modified
cap2		11/12/2010 06:54:27 PM
codingRegion_files		11/12/2010 07:12:24 PM
consen files		11/12/2010 07:12:24 PM
consensus_contigs.fas	15 KB	11/12/2010 07:11:56 PM
consensus_iter1_singletons.fas	1 KB	11/12/2010 07:04:04 PM
consensus_singletons.fas	4 KB	11/12/2010 07:11:56 PM
output		11/12/2010 07:12:11 PM

Done

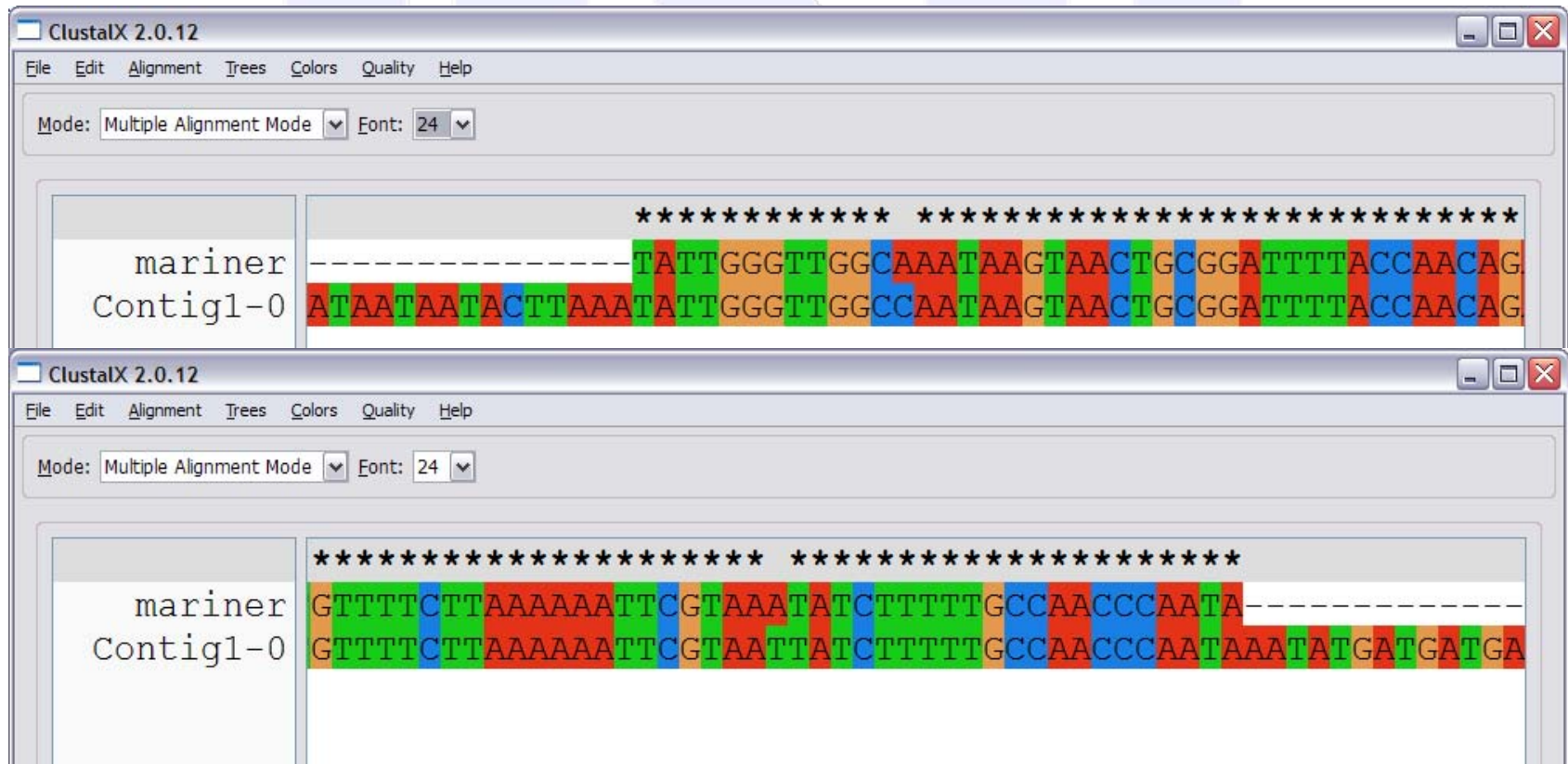
Y OF ME



TESeeker Results

□ Alignment

- TEsseeker top result with default parameters
- 99% identity with manually annotated *mariner*



TE Identification Summary

- ❑ Developed and automated a homology-based approach to identify TEs
 - Tedious and time-consuming task now automated
 - ❑ From months to hours or days
 - Output: high-quality consensus TEs
 - ❑ Can be used to determine instances in genome (density)
- ❑ Implemented as TESeeker
 - Distributed as a virtual appliance
 - ❑ All tools and scripts
 - Web interface
 - Distributed with high-quality library of representative coding regions from major TE families
- ❑ Approach contributed to multiple genome annotation projects
 - Sequences available in TEfam database
 - Most rigorously tested in arthropod genomes

Acknowledgments

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Questions or Comments?